

Pharmaceutical Nanotechnology

# Tumor-specific antibody-mediated targeted delivery of Doxil<sup>®</sup> reduces the manifestation of auricular erythema side effect in mice

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## Abstract

A mucocutaneous reaction in mice associated with Doxil<sup>®</sup> treatment was identified as auricular erythema (AE). Given that the immuno-targeting of Doxil<sup>®</sup> to tumors was found to influence also its systemic biodistribution pattern, the attempt was made to exploit a specific targeting of Doxil<sup>®</sup> to reduce the manifestation of this adverse reaction. This problem is of general significance, since cutaneous reactions often lead to alterations of Doxil<sup>®</sup> dosing regimen in patients and might subsequently compromise the therapeutic outcome of cancer treatment. Tumor-bearing mice were used to study the biodistribution and skin–tissue accumulation effects of the tumor-targeted Doxil<sup>®</sup> (the clinically used anti-cancer formulation) coupled with the anti-cancer monoclonal 2C5 antibody (mAb 2C5) as well as AE caused by Doxil<sup>®</sup> application. The modification of Doxil<sup>®</sup> with mAb 2C5 resulted in a significant decrease in the normal skin accumulation of doxorubicin compared to original Doxil<sup>®</sup> and substantially reduced AE. The frequency of AE was decreased by three to fourfold with the mAb 2C5-modified doxorubicin-loaded long-circulating liposomes. Thus, targeting of Doxil<sup>®</sup> with the anti-cancer mAb 2C5 not only can increase the tumor-specific accumulation of the drug, but also diminishes the cutaneous side effect of the original Doxil<sup>®</sup> therapy.

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## 1. Introduction

The enhanced tumoral delivery of anti-cancer drugs in long-circulating pharmaceutical nanocarriers is based on the passive accumulation of such carriers via the enhanced permeability and retention (EPR) effect responsible for the extravasation of large molecules and nanoparticles into the tumor mass in the cancerous areas with “leaky” vasculature (Iyer et al., 2006). Doxil<sup>®</sup> [doxorubicin-loaded polyethylene glycol(PEG)-coated liposomes] embodies this principle of passive targeting. It was shown earlier, that PEG-liposomes demonstrate prolonged circulation in the blood (Klibanov et al., 1990), and encapsulating various drugs including doxorubicin within these sterically

stabilized PEG-liposomes favorably alters their pharmacokinetics and biodistribution (Allen et al., 1991; Papahadjopoulos et al., 1991). This has resulted in the significant decrease in doxorubicin-associated dose-limiting toxicities: cardiomyopathy and myelosuppression (Berry et al., 1998; Safra et al., 2000).

However, due to this alteration of the pharmacokinetic profile of free doxorubicin, the dose-limiting toxicities of the long-circulating liposomal doxorubicin have been shifted towards mucocutaneous reactions such as palmar–plantar erythrodysesthesia (PPE) and mucositis/stomatitis (Hamilton et al., 2002; Lotem et al., 2000; Uziely et al., 1995). The current hypothesis for the development of PPE is that the small size and long circulation time of Doxil<sup>®</sup> allow for the liposome accumulation in the skin where the basal layers of the skin are damaged with prolonged exposure to doxorubicin as the liposomes slowly release this drug with known vesicant properties. Moreover, liposomes with long circulation times were found to accumulate in the skin of experimental animals to a greater extent than liposomes with shorter circulation times (Allen and Hansen, 1991; Papahadjopoulos et al., 1991). This is believed to be a result

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of the pressure-dependent extravasation of such liposomes into cutaneous tissues (Allen and Hansen, 1991; Hamilton et al., 2002; Hensley et al., 2001), which is currently supported by experimental and clinical data indicating that the accumulation of the long-circulating liposomes in fact repeats the anatomical distribution of PPE lesions in regions of skin that are subjected to the pressure or irritation, like the flexure creases of the hands, soles of the feet, or belt lines (Gordon et al., 2000; Hensley et al., 2001; Lotem et al., 2000). As indicated earlier, the incidence and severity of PPE associated with common Doxil<sup>®</sup> therapeutic regimens, by extrapolation of murine PK data to humans, can be controlled by increasing dose intervals, allowing time for the drug to clear from skin and for the ulcers to heal. Nevertheless, the benefit of dose delay would be offset by reduced therapeutic efficacy (Charrois and Allen, 2003a,b; Ranson et al., 1997; Uziely et al., 1995).

The modification of drug-loaded pharmaceutical carriers with tumor-specific ligands, such as anti-tumor monoclonal antibodies, is used to achieve higher local drug concentration at the target zone and enhanced anti-cancer effect of the chemotherapeutic agent (Noble et al., 2004; Park et al., 2004). The idea of adding monoclonal antibodies or their fragments to passively targeted long-circulating PEGylated drug carriers, including liposomes, may not only render the whole formulation more tumor-specific, hence demonstrating a superior therapeutic effectiveness (Torchilin, 2005), but could also result in decreasing drug presence in non-targeted areas, thus reducing possible adverse effects of drugs in long-circulating carriers onto normal areas of the body.

Earlier, we have described the monoclonal antinuclear antibody 2C5 (mAb 2C5) with the nucleosome-restricted specificity and unique ability to selectively bind to the surface of a broad variety of lymphoid and non-lymphoid tumor cells of murine and human origin (but not to the surface of normal cells) via the tumor cell surface-bound nucleosomes released from the apoptotically dying neighboring tumor cells (Iakoubov et al., 1995; Iakoubov and Torchilin, 1997). We have also used this antibody as a targeting moiety to actively target various drug-loaded long-circulating pharmaceutical nanocarriers, including doxorubicin-loaded PEGylated liposomes (Doxil<sup>®</sup>) to different tumors (Elbayoumi et al., 2007; Elbayoumi and Torchilin, 2006, 2007; Gupta et al., 2005; Lukyanov et al., 2004).

Here, we report our findings from the experiments with animals receiving different Doxil<sup>®</sup> formulations, plain and immuno-modified with the mAb 2C5. The animals subjected to original Doxil<sup>®</sup> treatment exhibited a blistering skin reaction, limited to the auricular region of the ears, hence named “auricular erythema” (AE). The manifestations of this AE cutaneous reaction closely resemble the PPE noticed earlier known for doxorubicin in PEG-liposomes. The occurrence and severity of the AE can be correlated with the reported drug release from the liposomes and its skin accumulation profiles. The data also suggest that the modification of Doxil<sup>®</sup> with the tumor-specific mAb 2C5 significantly reduces the manifestation of AE. Therefore, mAb 2C5-modification of Doxil<sup>®</sup> can potentially help alleviate the therapy-associated mucocutaneous adverse effects known for the original Doxil<sup>®</sup>.

## 2. Materials and methods

### 2.1. Materials

Doxil<sup>®</sup> was purchased from Pharmaceutics Inc. (West Roxbury, MA). Cholesterol (Chol), fully hydrogenated soy phosphatidylcholine (HSPC), *N*-(carbonyl-methoxy poly(ethylene glycol 2000)-1,2-distearoyl-*sn*-glycero-3-phosphoethanolamine sodium salt (MPEG2000-DSPE), and phosphatidylethanolamine (PE) were from Avanti Polar Lipids, Inc. (Alabaster, AL) and used without further purification. Doxorubicin was purchased from Sigma Chem., Inc. (St. Louis, MO). Triethylamine (TEA), octyl glycoside (OG) and diethylene triamine pentaacetic acid anhydride (DTPA) were also products of Sigma. Poly-oxethylene3400-bis(*p*-nitrophenyl carbonate) [PEG(*p*NP)<sub>2</sub>] was purchased from SunBio (Orinda, CA). Cell culture media, DMEM, EMEM and RPMI 1640, fetal bovine serum (FBS) and concentrated solutions of Na-pyruvate, non-essential amino acids, L-glutamine and penicillin/streptomycin stock solutions were purchased from CellGro (Kansas City, MO). Female BALB/C and C57/BL mice were purchased from Charles River Laboratories (Cambridge, MA). All solvents and other chemicals were analytical grade preparations.

Monoclonal 2C5 antibody was produced in ascites via the i.p injection of  $1.5 \times 10^6$  cells of the hybridoma cell line (obtained in our laboratory) into pristine primed BALB/C 4-week-old male mice. The production and the purification of the mAb 2C5 were carried out by Harlan Bioproducts (Indianapolis, IL).

### 2.2. Methods

#### 2.2.1. Antibody modification

To prepare mAb 2C5 conjugates with PEG3400-PE, a 40 molar excess of *p*-nitrophenylcarbonyl(*p*NP)-PEG-PE prepared as in (Torchilin et al., 2001), dispersed in 10 mg/ml solution of OG in 5 mM Na-citrate, 150 mM NaCl, pH 5.0, was added to an equal volume of 16.7 μM of the antibody in Tris-buffered saline (TBS), pH 8.5. The mixture was incubated for 24 h at pH 8.5 at 4 °C (Lukyanov et al., 2004).

#### 2.2.2. Preparation of antibody-modified liposomes

To modify doxorubicin-loaded PEG-liposomes, the mAb 2C5-PEG3400-PE conjugate obtained as described above was co-incubated with Doxil<sup>®</sup> preparation. For this purpose, the reaction mixture from the conjugation reaction (see the previous paragraph) was mixed in equal volumes with Doxil<sup>®</sup> and incubated for 6–8 h at 4 °C. Following the incubation, the remaining impurities and non-incorporated mAb 2C5 were removed by dialysis using cellulose ester dialysis tubes with a cutoff size of 250,000 Da. The complete removal of the free antibody was confirmed by the presence of only one peak of liposomes on the gel-chromatogram of the final sample (with no peaks corresponding to free mAb 2C5-PEG-PE or mAb 2C5-PEG-PE micelles) (Elbayoumi and Torchilin, 2007; Lukyanov et al., 2004).

### 2.2.3. Preparation of control liposomes

Control PEGylated liposomes mimicking Doxil<sup>®</sup> composition but containing no doxorubicin were prepared using the same lipid components and in the same concentrations as in Doxil<sup>®</sup>. A lipid film was obtained from MPEG2000-DSPE (3.19 mg/ml), HSPC (9.58 mg/ml), and Chol (3.19 mg/ml). The lipid film was suspended in HEPES-buffered saline (HBS), pH 7.4, and sonicated with a probe-type sonicator at 10 W power for 15 min, followed by several passages through the mini-extruder with 100 nm pore size polycarbonate filter, until approximately 100 nm liposomes with narrow size distribution were obtained (Elbayoumi and Torchilin, 2007; Lukyanov et al., 2004).

### 2.2.4. *In vitro* release of doxorubicin from liposomes

The *in vitro* release of doxorubicin from different Doxil<sup>®</sup> formulations was conducted in DMEM cell culture medium containing 10% fetal bovine serum (FBS). Liposomes at a concentration of 0.5 mg/ml of doxorubicin, diluted in the media, were sealed into dialysis tubes with the cutoff size of 12,000–14,000 Da. The liposome-loaded dialysis tubes were incubated in 50 ml of the media for 48 h at 37 °C, with continuous stirring. At various time points, aliquots were withdrawn, and replaced with the equal volume of the media. Doxorubicin concentrations were measured at 485 nm using a Hitachi U-1500 spectrophotometer, Hitachi Instruments (Schaumburg, IL) (Ishida et al., 2001; Xiong et al., 2005).

### 2.2.5. <sup>111</sup>In radiolabeling of liposomes

Doxil<sup>®</sup>-mimicking liposomes containing the amphiphilic chelate DTPA-PE (HSPC:Chol:MPEG200-DSPE:DTPA-PE in 3:2:0.3:0.3 molar ratio) were prepared along with the mAb 2C5-immunoanalogues. The loading of the liposome-incorporated DTPA-PE with <sup>111</sup>In was performed via the transchelation mechanism from a weak citrate complex. DTPA-PE-containing liposomes were supplemented with 0.1 M citrate buffer and incubated for 1 h with <sup>111</sup>In (as <sup>111</sup>In chloride in citrate buffer) at RT, and then dialyzed overnight against HBS at 4 °C to remove the free label (Elbayoumi and Torchilin, 2006; Torchilin et al., 2001).

### 2.2.6. Growth of tumors in mice

Murine breast carcinoma (4T1) and colon carcinoma (C26) tumors were implanted in 8-week-old BALB/C mice by the subcutaneous injection of 10<sup>5</sup> cancer cells into the fat pads in the lower abdominal region. Similarly, murine Lewis lung carcinoma (LLC) cells were S.C. injected in 8-week-old C57/BL mice (5 × 10<sup>4</sup> cells/mouse). The time for the appearance of the palpable tumor (varies from one cell line to another and usually is 7–12 days). Mice were regularly monitored, with free access to food and water (following animal care protocol no. R01210 approved by Northeastern University Institutional Animal Care and Use Committee, in accordance with the “Principles of laboratory animal care”, NIH publication no. 85–23, revised in 1985), and tumor volumes were calculated by measuring the length and width of the tumor at regular intervals (Chakilam et al., 2004; Elbayoumi and Torchilin, 2006).

### 2.2.7. Single-dose pharmacokinetics and biodistribution of <sup>111</sup>In-labeled liposomes

*In vivo* biodistribution studies of <sup>111</sup>In-radiolabeled Doxil<sup>®</sup>-mimicking liposomal formulation along with mAb 2C5-modified analogue were performed in 8-week-old female BALB/C mice, both healthy and 4T1 tumor-bearing (tumor volume range: 200–300 mm<sup>3</sup>), in two separate experiments. In each experiment, BALB/C mice were injected with 0.1 ml of 1 mg/ml <sup>111</sup>In-radiolabeled Doxil<sup>®</sup>-mimicking liposomes via the tail vein. At 15, 30, 120, 360, 720 and 1440 min postinjection, blood was collected using a Pasteur pipette from the retro-orbital plexus of the eye, and the mice were euthanized by CO<sub>2</sub> followed by collection of different organ tissues. The amount of radioactivity was quantified as CPM using a Beckman 5500B gamma-counter, and the amount of the accumulated radioactivity per gram of tissue was calculated followed by calculation of the temporal biodistribution and tissue accumulation parameters (Chakilam et al., 2004; Elbayoumi and Torchilin, 2006; Pastorino et al., 2003).

### 2.2.8. Assessment of auricular erythema as drug-induced side effect

Starting 1 week after the initiation of the drug administration (given in a sub-therapeutic dose regimen, as 2.2 mg/kg dose every 5 days, for four consecutive doses) into tumor-bearing mice, location, count and severity of any developed auricular erythema (AE) lesions were monitored and documented for the rest of the treatment, in BALB/C mice bearing C26 tumors, and C57/BL bearing LLC tumors. Finally, AE percentage (AE%) was estimated for each group.

### 2.2.9. Statistics

Differences in the apparent AE occurrence percentage, and percentage of ears exhibiting AE manifestations during the treatment were compared using logistic regression, and comparing the odds ratio (OR) between the treatment groups [(AE events vs. no event) of mAb-2C5-modified Doxil<sup>®</sup> intervention/(AE events vs. no event) of original Doxil<sup>®</sup> intervention]. The significance level for the OR proportion of probability (OR<sub>p</sub>) was set at 0.1 (Batist et al., 2006; Greenhalgh, 1997; Potamianou et al., 2005).

## 3. Results and discussion

### 3.1. Doxorubicin leakage from antibody-modified liposomes

Similar to earlier data (Elbayoumi and Torchilin, 2006, 2007; Lukyanov et al., 2004), the post-insertional technique for immuno-modification of Doxil<sup>®</sup> with the mAb 2C5-PEG-PE conjugate did not markedly alter the physicochemical properties of the original Doxil<sup>®</sup> formulation and resulted in the final preparation of 112 ± 7 nm liposomes with 73 ± 3 antibody molecules attached per single liposome. The complete release profiles (Fig. 1, panel A) of the original Doxil<sup>®</sup> and mAb 2C5-Doxil<sup>®</sup> also did not show any significant difference between these formulations. These *in vitro* release profiles have also

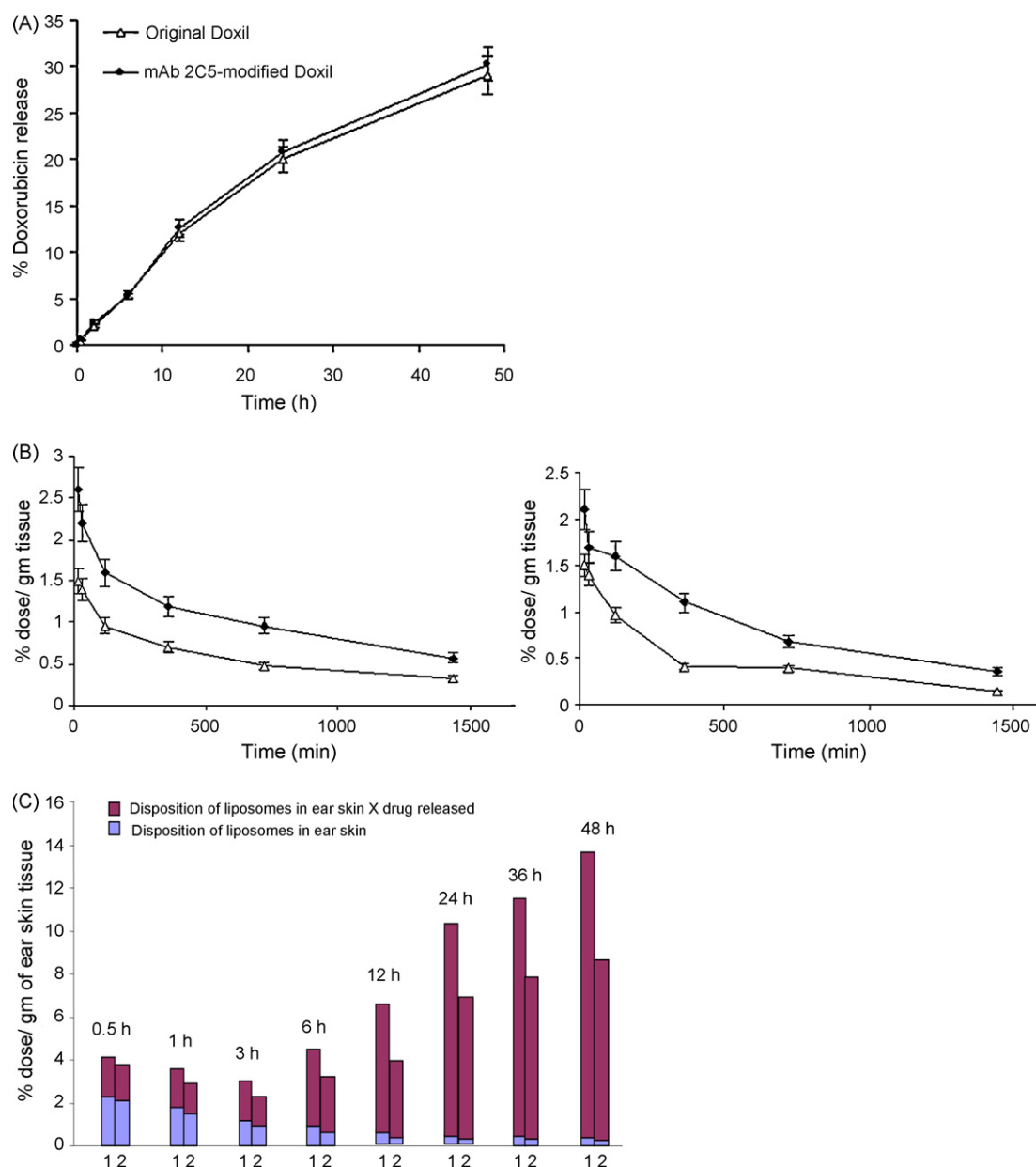


Fig. 1. (A) *In vitro* doxorubicin release from various Doxil® preparations in cell culture media containing 10% BSA, (B) biodistribution data of  $^{111}\text{In}$ -labeled Doxil®-mimicking liposomal preparations in skin (auricular) tissue of the ears, in healthy BALB/C mice (left panel); and 4T1-tumor-bearing mice (right panel), (◆) Doxil®-mimicking liposomes; and (△) mAb 2C5-modified Doxil®-mimicking liposomes ( $n=4$ , results indicated  $\pm$ S.D.) and (C) hybrid representation of the different Doxil®-mimicking liposome accumulation and doxorubicin release profiles; 1-original Doxil®, 2-mAb 2C5-Doxil®, determined for 24 h and extrapolated up to 4 days.

demonstrated a slow first order release of doxorubicin from its carrier with just about 30% drug release after 2 days, which extrapolates to ca. 80% of the encapsulated drug to be released after approximately 5 days. This corresponds well to the estimated *in vitro* doxorubicin release half-time of 85 h for similar HSPC liposomes (Allen et al., 2005). This noteworthy prolonged release profile of the drug from the liposomal carrier, extending for up to five or even more days, evidently plays an important part of sustaining an effective local tissue concentration at the tumor site, combined with an extended doxorubicin tumor half-life reaching up to 70 h, as demonstrated earlier by other groups (Charrois and Allen, 2003a,b, 2004; Papahadjopoulos

et al., 1991). On the other hand, it may play a causative role in the necrotic manifestations at normal mucocutaneous tissues, especially on the multiple dosing treatment regimen of Doxil®. Especially when considering the reported protracted skin-tissue half-life (approximately 80–110 h) of the released drug (Charrois and Allen, 2003a,b, 2004).

### 3.2. *In vivo* biodistribution and skin accumulation of $^{111}\text{In}$ -labeled liposomes

Using liposomes mimicking the lipid composition of Doxil® and radiolabeled with  $^{111}\text{In}$ , the tissue biodistribution of dif-

ferent liposomal formulations was determined. Blood clearance data of analogous formulations in tumor-bearing BALB/C mice reported earlier (Elbayoumi and Torchilin, 2006), indicated that mAb 2C5-Doxil<sup>®</sup>-mimicking liposomes cleared faster from the body compared to plain liposomes (MRT  $\approx$  12 h for mAb 2C5-liposomes, compared to approximately 17 h for plain liposomes, as derived from AUC data using a non-compartmental analysis model). This somewhat accelerated clearance of the antibody-modified liposomes from the body can be attributed to the presence of antibody molecules on the liposomal surface, which made liposomes more prone to the uptake by RES cells, and this is true for the cases when both tumor-specific and non-specific immunoglobulins are present on the PEG-liposome surface (Chakilam et al., 2004; Xiong et al., 2005). However, mAb 2C5-immunoliposomes still demonstrated a sufficiently prolonged circulation, which corresponds well to the results reported with similar formulations in healthy mice (Elbayoumi and Torchilin, 2006; Pastorino et al., 2003). Furthermore, one can also expect that the higher dose of liposomes ends in the tumor in the case of mAb 2C5-Doxil<sup>®</sup> (Elbayoumi and Torchilin, 2006).

As for the accumulation in the normal skin–tissue of the ears (Fig. 1, Panel B), it is lower for mAb 2C5-immunoliposomes compared to original Doxil<sup>®</sup>, most probably due to their shorter blood residence time. Further more, one can suggest that in case of tumor-bearing mice, the tumor-targeted nature of immunoliposomes could be partially involved, when the tumor attracts and accumulates circulating targeted liposomes in a shorter time period (revealed as a shorter plasma half-life), similar to what was described in earlier reports (Charrois and Allen, 2003a,b; Elbayoumi and Torchilin, 2006).

Therefore, combining the data collected from both the liposomal drug release profiles with the skin accumulation profiles clearly reveals an important fact. Despite the minor differences in skin accumulation between different liposomal preparations, especially at longer post-injection times, a significant amount of doxorubicin estimated to be released in the same time period would result in a marked amplification of these differences of the liposomal skin accumulation. Fig. 1, panel C, illustrates the differential doxorubicin accumulation in the ear area, produced by factoring the liposomal accumulation by doxorubicin release. The graph clearly demonstrates that even after 48 h post-injection (extrapolated), according to the drug release profile, the amount of doxorubicin accumulated in the ear tissue areas in case of mAb 2C5-Doxil<sup>®</sup> is still significantly lower than for the original Doxil<sup>®</sup>.

### 3.3. Assessment of drug-associated auricular erythema as toxicity-indicative side effect

In accordance with the theory of pressure-dependent extravasation of PEG-liposomes and PPE (Charrois and Allen, 2003a,b; Lotem et al., 2000), we observed that therapy-related symptoms appeared in mice treated with the different Doxil<sup>®</sup> formulations to a variable extent (but not in the PBS control groups). Observed skin damages are similar to the PPE lesions, but more confined to the auricles of the ears and tend to cause serious blistering and

desquamation of the auricular tissue (Fig. 2, Panels A and B), hence termed “auricular erythema (AE)”. It was first observed in C57/BL mice starting after the administration of the second Doxil<sup>®</sup> dose (2.2 mg/(kg q 5 days)), and in the BALB/C mice after the administration of the third Doxil<sup>®</sup> dose. Tables 1 and 2 show the data collected after recording the incidence of this AE side effect during the entire period of therapy.

A plausible explanation for the appearance of AE lesions relates to the fact that the external auricles in mice ears have a relatively thin cutaneous tissue (acting as a cooling system to allow for the efficient dissipation of the body heat). Hence, the slow release of doxorubicin from the PEG-liposomes accumulated in the skin of the ear after several doses, causes severe necrosis in surrounding tissue. We have used this AE side effect as a means of estimating the toxicity related to the treatments with different Doxil<sup>®</sup> preparations.

It is clearly seen that the number of C57/BL mice exhibiting this AE side effect is noticeably lower in the case of mAb 2C5-Doxil<sup>®</sup> (ORp  $\leq$  0.1) than in the case of the original Doxil<sup>®</sup> group. A similar pattern was also noted in the case of BALB/C mice. There is a clear significant difference in AE percentage between the original Doxil<sup>®</sup> treatment group and the mAb 2C5-Doxil<sup>®</sup> group. This low incidence of AE in case of animals treated with mAb 2C5-Doxil<sup>®</sup> compared to that of the original Doxil<sup>®</sup> formulation can be related to both the shorter blood-residence time of this immuno-formulation and to its tumor-specific accumulation. It seems that tumor-specific ligands, such as mAb 2C5, might play a considerable role in lowering side effects of Doxil<sup>®</sup>, by actively limiting its presence to the tumor tissue, hence avoiding non-targeted tissues and related side effects. Nevertheless, further toxicological and pharmacokinetic studies concerning the AE and PPE phenomena, using larger number of animals/treatment group are still needed, in order to establish a rigorous correlation between the incidence profiles of AE in the animals and the dose intensities of different Doxil<sup>®</sup> formulations.

Table 1  
Occurrence of auricular erythema in LLC-bearing C57/BL mice, corresponding to each treatment

Treatment	% AE (No. C57/BL mice with AE/gp)	% of ears with AE (no. of ears with AE/gp)
PBS	0% (0/10)	0% (0/20)
Doxil <sup>®</sup>	80% (8/10)*	80% (16/20)*
mAb-2C5-Modified Doxil <sup>®</sup>	30% (3/10)*	20% (4/20)*

\* $P \leq 0.1$

Table 2  
Occurrence of auricular erythema in C26-bearing BALB/C mice, corresponding to each treatment

Treatment	% AE (no. of BALB/C mice with AE/gp)	% of ears with AE (no. of ears with AE/gp)
PBS	0% (0/10)	0% (0/20)
Doxil <sup>®</sup>	56% (5/9)	56% (10/18) <sup>†</sup>
mAb-2C5-modified Doxil <sup>®</sup>	20% (2/10)	10% (2/20) <sup>†</sup>

<sup>†</sup>  $P \leq 0.1$

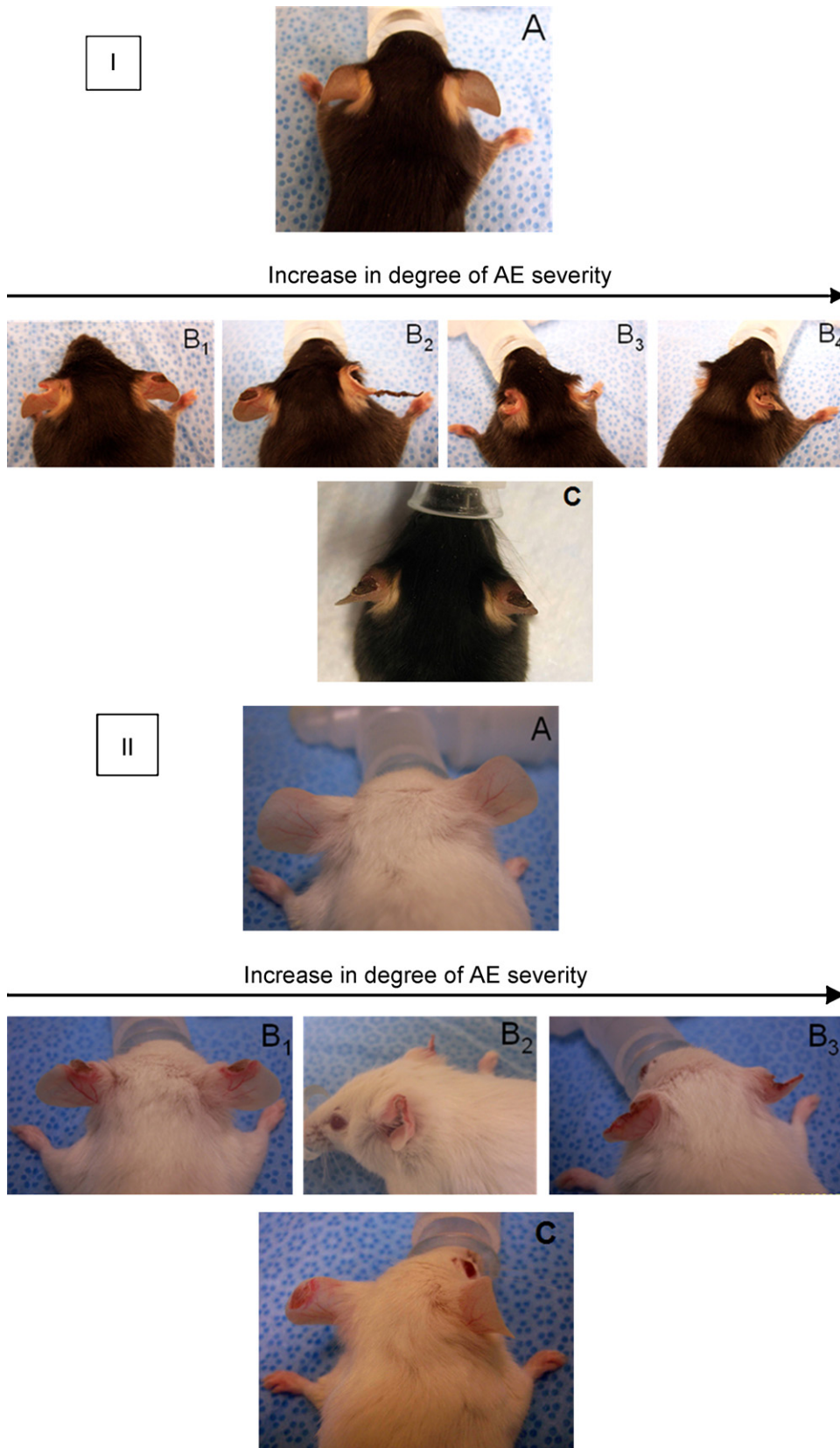


Fig. 2. Auricular erythema manifestation in C57/BL mice implanted with LLC tumors (I); and BALB/C mice implanted with C26 tumors (II), as a result of drug administration, showing untreated mouse (A); compared to Doxil<sup>®</sup>-treated mice (B<sub>1-4</sub>), exhibiting the progression of AE manifestation during chemotherapy; and to mAb 2C5-Doxil<sup>®</sup>-treated mice (C, showing AE manifestation developed only after the last administered dose).

#### 4. Conclusions

Here, we report a new cutaneous side effect of long-circulating liposomal carriers encapsulating the anti-cancer drug doxorubicin observed in mice. This syndrome appearing in the ear skin areas and termed “auricular erythema”, is associated with Doxil® treatment, hence becomes a candidate to be added to the family of the mucocutaneous adverse reactions caused by the long-circulating liposomal doxorubicin.

Our data suggest also that the immuno-modification of Doxil® with the anti-tumor antibody, mAb 2C5 in this particular case, can create a formulation that possesses the ability to specifically recognize and accumulate in various tumors, and at the same time lowers its presence in normal tissues. Therefore, the active targeting of the long-circulating liposomal preparations of doxorubicin, for example, with the mAb 2C5, presents an attractive opportunity to further control the *in vivo* behavior of the original Doxil® and improve its tolerability.

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